

Authors' Reply

Authors' reply to 'Granulocytic myeloid-derived suppressor cells in peripheral blood of patients with cutaneous melanoma'

We are grateful to Dr Franklin and Dr Schilling for their interest in our recent article (1) and we will try to respond to their constructive critical review.

In regard to Dr Franklin's and Dr Schilling's emphasis that the CD10⁻CD15⁺CD14⁻HLA-DR^{-/low}CD33^{low}CD11b⁺CD45^{low}CD16^{low}Lin⁻ panel is '... a widely used panel for identifying grMDSC in PBMC after density centrifugation', we would like to underline that we initially conducted a pilot study showing that cells with the above-mentioned phenotype copurified with the PBMC on density gradients (Supplementary Figure 1, available at *International Immunology Online*). The reasons why we decided to continue the study with lysed peripheral blood samples were listed in the article (1). We agree that the activated neutrophils can change their buoyancy and copurify on a density gradient, but these activated cells are still CD10⁺ and the CD10 expression could even be up-regulated in activated mature neutrophils (2).

Regarding the remark on the separation of the grMDSC from eosinophils, besides being CD10⁻, eosinophils are also CD16⁻, CD45^{bright} and high on side scatter (SSc) (3). Our targeted population showed low, but positive CD16 expression, low CD45 expression and a low SSc position (1). Additionally, no eosinophilia was recognized in differential blood counts in our patients with >10% of the presumed grMDSC population. Accordingly, we disagree with Dr Franklin's and Dr Schilling's assertion that we have distinguished eosinophils and grMDSC on the basis solely of low or absent CD16 expression.

We agree that antigen expression changes with aging; however, both the grMDSC frequencies and the CD16 expression intensity did not correlate with the patient's age in our study. Subsequently, we detected CD66b on our grMDSC, which could separate them from immature myeloid cells, according to the results of Dumitru's group, cited by Dr Franklin and Dr Schilling.

We agree that the functional assay is the most reliable evidence for immunosuppressive activity of certain cell populations; thus, we titled our manuscript as 'A subpopulation that may correspond to ...' (1). On the other side, the classical suppressive assay can exclude neither the influence of activated healthy granulocytes, which are also capable of T-cell suppression (4), nor the possibility of MDSC activation during long-time cultivation with T cells (5). The suppressive assay is certainly superior in the situation of testing MDSC from the site of inflammation or the tumor-growing site, because those cells possess the immediate capacity to down-regulate T-cell proliferation *in vitro*

(5). Instead of using the classical suppressive assay, we identified our grMDSC by phenotype (1) and investigated possible *in vivo* grMDSC suppressive outcomes related to arginine depletion, oxidative stress and immune response skewing (Supplementary Figure 2, available at *International Immunology Online*).

At the end, we consider as a weakness of our study the fact that we have not performed repeated measurements of the grMDSC frequencies during the follow-up period.

Supplementary data

Supplementary data are available at *International Immunology Online*.

Ivan Stanojevic
Department of Clinical and Experimental Immunology,
Institute for Medical Research, Military Medical Academy,
Cnotravaska 17, 11000 Belgrade, Serbia

Mirjana Djukic
Department of Toxicology, Faculty of Pharmacy,
University of Belgrade, 11000 Belgrade, Serbia

Danilo Vojvodic
Department of Clinical and Experimental Immunology,
Institute for Medical Research, Military Medical Academy,
Cnotravaska 17, 11000 Belgrade, Serbia

Correspondence to: I. Stanojevic;
E-mail: ivanivanstanojevic@gmail.com

References

- 1 Stanojevic, I., Miller, K., Kandolf-Sekulovic L. *et al.* 2016. A subpopulation that may correspond to granulocytic myeloid-derived suppressor cells reflects the clinical stage and progression of cutaneous melanoma. *Int. Immunol.*, 28:87–97. doi:10.1093/intimm/dxv053.
- 2 Shipp, M. A., Stefano, G. B., Switzer, S. N., Griffin, J. D. and Reinherz, E. L. 1991. CD10 (CALLA)/neutral endopeptidase 24.11 modulates inflammatory peptide-induced changes in neutrophil morphology, migration, and adhesion proteins and is itself regulated by neutrophil activation. *Blood* 78:1834.
- 3 Gorczyca, W., Sun, Z. Y., Cronin, W., Li, X., Mau, S. and Tugulea, S. 2011. Immunophenotypic pattern of myeloid populations by flow cytometry analysis. *Methods Cell Biol.* 103:221.
- 4 Schmielau, J. and Finn, O. J. 2001. Activated granulocytes and granulocyte-derived hydrogen peroxide are the underlying mechanism of suppression of t-cell function in advanced cancer patients. *Cancer Res.* 61:4756.
- 5 Haverkamp, J. M., Crist, S. A., Elzey, B. D., Cimen, C. and Ratliff, T. L. 2011. *In vivo* suppressive function of myeloid-derived suppressor cells is limited to the inflammatory site. *Eur. J. Immunol.* 41:749.